

## Solid-state $^{14}\text{N}$ NMR of amino acids and polypeptides

Stanislav L. Veinberg,<sup>a</sup> Kristopher J. Harris,<sup>a</sup> Luke A. O'Dell<sup>b</sup> and Robert W. Schurko<sup>a</sup>

(a) Department of Chemistry and Biochemistry, University of Windsor, Windsor, Ontario

(b) Steacie Institute for Molecular Sciences, National Research Council, Ottawa, Ontario

[rschurko@uwindsor.ca](mailto:rschurko@uwindsor.ca)

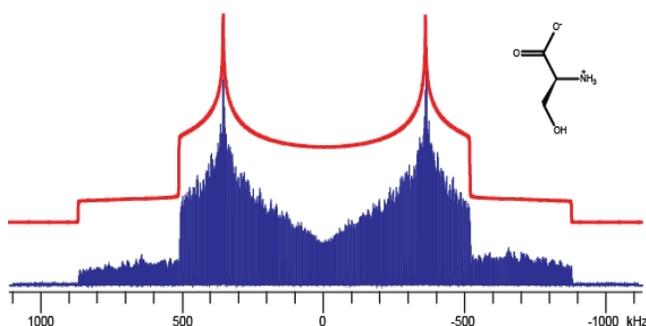
Amino acids play a vital role as the building blocks of proteins and their arrangement in the polypeptide chain controls protein structure and folding. These systems have been extensively investigated by NMR using  $^{13}\text{C}$  and  $^{15}\text{N}$  as the probe nuclei. Unfortunately, due to the low natural abundances of the probe nuclei (especially of  $^{15}\text{N}$ ), costly isotopic labelling is often necessary to obtain high quality spectra. Nitrogen has two NMR active nuclei ( $^{14}\text{N}$  and  $^{15}\text{N}$ ), the former is 99.63% naturally abundant and perhaps may have use as an alternative probe nucleus, despite difficulties associated with  $^{14}\text{N}$  NMR.

Nitrogen-14 is an integer-spin nucleus ( $I = 1$ ) with a non-zero electric quadrupole moment ( $eQ = 20.44$  mb) and a low gyromagnetic ratio ( $\gamma = 1.93 \times 10^7$  rad  $\text{T}^{-1} \text{s}^{-1}$ ) [1]. Due to the quadrupolar nature of this nucleus, it couples to the surrounding electric field gradient (EFG), which is dependent upon the local ground state electronic environment. Solid-state  $^{14}\text{N}$  NMR spectra of micro-crystalline samples often yield powder patterns with widths exceeding 1 MHz [1]. Spectral broadening low Larmor frequencies make acquisitions of  $^{14}\text{N}$  powder patterns quite difficult.

Utilizing the 21.1 T NMR spectrometer and the WURST-QCPMG pulse sequence [2,3], we have been able to acquire high quality powder patterns of the majority of known amino acids with generally

short acquisition times. In particular, we have found that pseudo-tetrahedral nitrogen environments,  $\text{R-NH}_3^+$  (L-serine and L-threonine) and  $\text{R}_2\text{-NH}_2^+$  (*trans*-4-hydroxy-L-proline), are amenable to  $^{14}\text{N}$  NMR experiments.

Pseudo-tetrahedral nitrogen environments have quadrupolar coupling constant ( $C_Q$ ) values ranging from 0.8 to 2 MHz, and can't be differentiated based on  $C_Q$  alone, however, there is a clear variation in the asymmetry



**Figure 1:**  $^{14}\text{N}$  powder pattern of L-serine (blue) at 21.1 T. This spectrum is composed of 4 sub-spectra and took *ca.* 1.5 hours to acquire. Spectral simulation (red) yields  $C_Q = 1.165$  MHz and  $\eta_Q = 0.18$ .

parameter ( $\eta_Q$ ). We have found that pseudo-tetrahedral nitrogen environments may be grouped into low (0.0 – 0.25), intermediate (0.26 – 0.6), and high (0.61 – 1.0)  $\eta_Q$  classes.

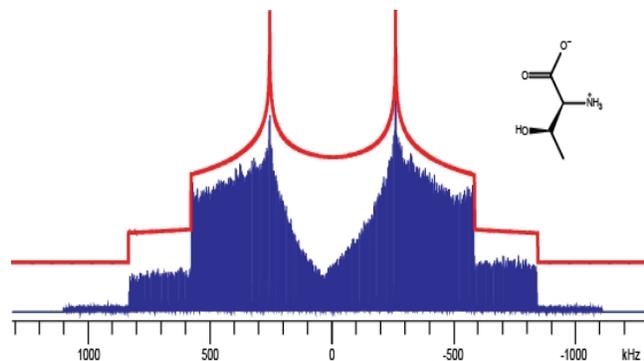
The sensitivity of the  $^{14}\text{N}$  NMR spectrum to surrounding EFG is effectively demonstrated with three representative examples. Figure 1 depicts the  $^{14}\text{N}$  powder pattern of L-serine ( $\eta_Q = 0.18$ ). This type of spectrum is characteristic of  $\text{C-NH}_3^+$  environments which have cylindrical pseudo- $C_3$  symmetry about the C-N bond due to absence of intermolecular interactions.

Figure 2 depicts the  $^{14}\text{N}$  powder pattern of L-threonine ( $\eta_Q = 0.385$ ). The intermediate  $\eta_Q$  values arise because of varying degrees of intermolecular interactions which cause deviation from cylindrical symmetry about the C-N bond.

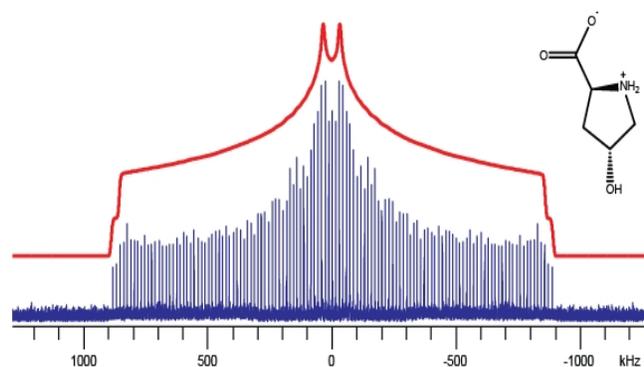
A  $^{14}\text{N}$  powder pattern of *trans*-4-hydroxy-L-proline ( $\eta_Q = 0.93$ ) is depicted in Figure 3. High  $\eta_Q$  values are characteristic of an entirely different type of nitrogen environment, with two protons and two carbon atoms bound to the nitrogen, and no pseudo- $C_3$  rotational axis.  $^{14}\text{N}$  EFG tensor calculations using plane-wave DFT methods (i.e., CASTEP [4]) are in good agreement with experimental findings. Further investigations of the remaining amino acids are underway. We are expanding this study to amino acids containing side-group nitrogen atoms (histidine, lysine, arginine, etc.) as well as larger peptide systems (di- and tripeptides).

#### References

- [1] P. Pykkö, *Mol. Phys.* **99** (2001) 1617.
- [2] L.A. O'Dell and R.W. Schurko, *Chem. Phys. Lett.* **464** (2008) 97.
- [3] L.A. O'Dell and R.W. Schurko, *J. Am. Chem. Soc.* **131** (2009) 6658.
- [4] S.J. Clark, M.D. Segall, C.J. Pickard et al. *Z. Kristallogr.* **220** (2005) 567.



**Figure 2:**  $^{14}\text{N}$  powder pattern of L-threonine (blue) at 21.1 T. This spectrum is composed of 4 sub-spectra and took ca. 2 hours to acquire. Spectral simulation (red) yields  $C_Q = 1.12$  MHz and  $\eta_Q = 0.385$ .



**Figure 3:**  $^{14}\text{N}$  powder pattern of *trans*-4-hydroxy-L-proline (blue) at 21.1 T. This spectrum is composed of 2 sub-spectra mirrored about 0 ppm and took ca. 3 hours to acquire. Spectral simulation (red) yields  $C_Q = 1.19$  MHz and  $\eta_Q = 0.93$ .